

Methods for the Collection and Characterization of Airborne Particles in the Textile Industry

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ABSTRACT

The paper presents the results of the determination of indoor and outdoor air concentration levels in textile companies, to identify the areas with the highest concentration level, by using an online recording system such as the Laser Aerosol Spectrometer MINI LAS model 11-E. The next step was the collection of fibers, namely micro and nano plastic particles (MNPs) from the vicinity of the workplaces of polyester, polyamide, and polypropylene fibers processing units in the textile industry in Romania, to obtain a sufficient quantity for laboratory analysis to determine the size and shape of the particles as well as their chemical composition. Two types of pumps were used, differentiated by their operating parameters: TECORA SKYPOST with air-flow of 38 l/min and GILAIRPLUS with airflow 2l/min. Filters made of different materials with different diameters and pore sizes were used. Several times more PM₁₀ than PM₁ (74.5 μg compared to 12.5 μg) was found. In all cases, both particles and fibers showed the same Raman fingerprint. The GS-MS analyses showed some contamination of the workspaces with particles other than the processed fibers. The most viable filters are Si filters with a pore size of 10 microns to 1 micron and the use of the selected collection and transport filter system

Keywords: Micro and nanoparticles, Concentration, Collection, Physical and structural analysis

INTRODUCTION

Airborne particulate matter as complex mixtures of organic and inorganic substances from different sources of particle emissions is one of the main air pollutants (Grosu et al., 2023). However, the abundance and varied composition of the suspended particulate matter make it difficult to locate the precise precursors for some of these environmental pollutants.

Depending on the emission source, particulate matter (PM) can vary in size / aerodynamic diameter (ultrafine PM: 0.01 μm – coarse PM: 100 μm),

density, shape, and composition (Qader et al., 2023). PM from both natural and anthropogenic sources is known to affect the environment (air, water, soil) and human health. Due to its microscopic size, PM can get deep into the lungs and bloodstreams, threatening human health, accounting for nearly one-third of deaths from chronic respiratory disease, lung cancer, and one-quarter of deaths from heart attacks (Agay-Shay et al., 2013). More, according to the latest OECD report, air pollution will be the largest environmental cause of premature death worldwide by 2050, unless collective action is taken (von Schneidemesser et al., 2015). As particular types of PM, plastic particles matter contamination and associated weathering mechanisms are emergent environmental and human food chain concerns (Klingelhofer et al., 2020). There are multiple sources of micro and nano plastic particles split into two main categories: *primary* (produced as microparticles or enters the natural environment already as microparticles) and *secondary* (generated by fragmentation or abrasion of other products in the natural environment) (Grosu et al., 2023). The textile industry, as a major polluter worldwide, generates micro and nano plastic particles during the manufacturing processes, especially nylon, polyester, polyurethane, polyolefin, acrylic, and vinyl-type polymers (Visileanu et al., 2023). Various reviews highlight synthetic textiles as the main superordinate source of airborne microplastics and fibers were the most abundant shape of MP found in the atmosphere (Schopel & Stamminger, 2019).

The paper presents the results of the identification, counting, collection, and characterization of micro and nanoparticles generated during various technological processes in the textile industry.

MATERIALS AND METHODS

Determining PM Concentration

In the first step, we had to select the workplace with the highest total suspended particles in the air, to demonstrate the presence of the plastic particle in the air and that is possible to collect such amount that is necessary for characterization. The level of concentration (total suspended particles-TSP, PM₁₀, PM_{2.5}, and PM ($\mu\text{g}/\text{m}^3$) and total counted particles-TC (l/l) in different work areas in the weaving and knitting mill that produce textile fabrics from synthetic fibers (polyester-PES/PET, polyamide-PA, and polypropylene-PP) were determined by using an online portable Grimm Optical Spectrometer MINI LAS model 11-E (Grimm Aerosol Technik, n.d.). The mini device uses high-tech laser optics to capture each particle with dimensions between 0.25 and 32 μm and classifies them into 31 size channels (via software), according to the European standard EN 481.

Sampling of MNPs

The collection of particles was performed by using 2 methods:

- Method 1 - according to SR EN 12341: 2014 standard which is based on sampling the particles on the filters and weighing them using a balance. The TECORA-SKYPOST PM-HV (Figure 1). type device was used for

sampling, which ensures the volumetric measurement with dry gas ether with an accuracy of $\pm 2\%$. Airflow 38 l/min, 6 m³/h rotary vane pump type. Quartz filters with a diameter of 47 mm were used (see Figure 2). The volume of air was recorded during the collection period. Particles were taken from each type of 3 filters (A1, A2, A3).

- Method II - is based on sampling the particles on the filters using a GILAIR PLUS - type device equipped with cyclone Higgins-Dewell (HD) and a 2 l/min air flow pump. For sampling, 37 mm diameter quartz filters (B1, B2, B3) and 25 mm diameter nucleopore filters coated with gold membrane (C1, C2, C3) were used. For each type of particle and filter, 3 filters were taken. The number of particles collected was determined by the difference in mass between the white filters and the filters after collection.



Figure 1: TECORA SKYPOST PM-HV sampling device.



Figure 2: GILAIR PLUS air sampling device.

Table 1 shows the values of the quantities of PES/PET, PA and PP particles collected, from which it can be seen that the highest quantity was obtained when using \varnothing 37mm quartz filters and GILAIR PLUS pump with 2 l/min airflow for PA particles.

Table 1. Particles quantity samples.

	Collection parameters\characteristics											
	PES/PET						Filter mass (μg)					
	Filter S1	Filter S2	Filter S3	Filter A1	Filter A2	Filter A3	Filter P1	Filter P2	Filter P3	PA	PP	
Quartz filter, $\varnothing 37\text{mm}$	88.32	87.66	86.78	87.69	87.70	92.33	92.34	87.01	87.81			
Initial mass, μg												
Mass after collection, μg	89.45	89.39	88.33	88.38	88.30	92.79	95.85	90.60	94.56			
Airflow: 2 l/min	1.13	1.72	1.54	683	601	462	3.51	3.58	6.74			
Mass of collected particles, μg												
Air concentration, $\mu\text{g}/\text{m}^3$	6.08	8.85	11.90	1.208	0.896	0.298	28.52	17.47	34.82			
Air volume, l	172.3	185.2	122.5	247.42	242.11	261.6	120.07	200.48	191.30			
Polycarbonate with	9.101	9.393	9.463	9.014	9.569	9.077	9.489	9.9925	9.762			
Initial mass, μg												
Mass after collection, μg	9.124	9.409	9.487	9.030	9.579	9.094	9.511	9.952	9.777			
Mass of collected particles, μg	23.0	16.0	24.0	16.0	10.0	17.0	22.0	27.0	15.0			
Au-coated membrane, $\varnothing 25\text{mm}$	135.4	90.20	119.21	117.41	68.51	123.43	78.82	127.47	114.37			
Air concentration, $\mu\text{g}/\text{m}^3$												
Air volume, l	1669	1774	2013	136,3	146,0	137,7	279,1	211,8	131,2			
Air flow-2 l/m Temperature, $^{\circ}\text{C}$					19.3							

EXPERIMENTAL

The database created as a result of the records made for each fraction was populated with at least 50 values for each variable. The main statistical indicators for this variable were obtained by using a specialized software program.

The mean, dispersion and standard deviation, median and quartiles, skewness, and kurtosis were calculated for each of the four variables considered to define air quality in the vicinity of the workplaces, to highlight cases where action should be taken. Histograms and box plots were plotted for TSP, PM10, PM2.5, and PM.

The comparative results (indoor/outdoor) for the companies processing PES/PET fibers are shown in Figure 3. The highest value of PES/PET particle concentration was recorded inside the company processing recoverable PET materials ($127.5 \mu\text{g}/\text{m}^3$); this value is about 4 times higher than the one recorded outside ($36.7 \mu\text{g}/\text{m}^3$).

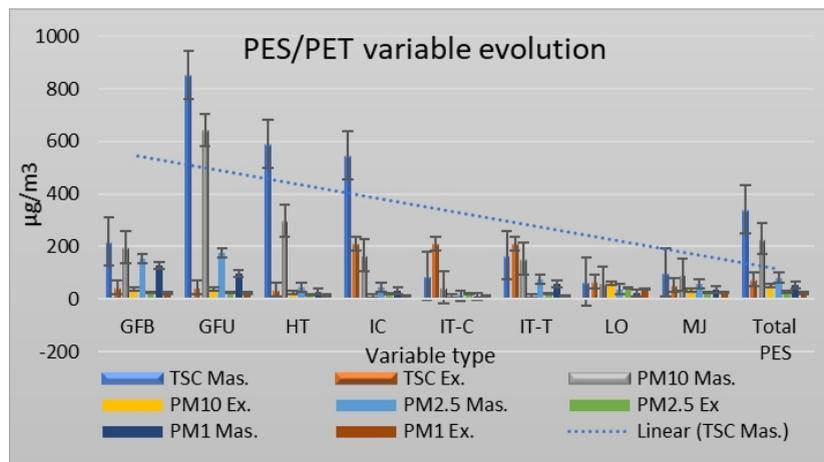


Figure 3: PES/PET PM concentration level comparison (indoor/outdoor).

Figure 4 shows the comparative results (indoor/outdoor) for companies processing PA fibers. In this case, the highest value of PM1 concentration of polyamide particles recorded in the vicinity of workplaces is $81.2 \mu\text{g}/\text{m}^3$ which is approx. 7 times higher compared to the outdoor value of $11.86 \mu\text{g}/\text{m}^3$. The average PM1 concentration recorded at the two companies is $71.57 \mu\text{g}/\text{m}^3$. The highest value of polyamide PM1 concentration recorded in the vicinity of the workplaces was obtained at MG company ($81.2 \mu\text{g}/\text{m}^3$). The outdoor PM 1 value was $11.86 \mu\text{g}/\text{m}^3$. Figure 5 shows the comparative results (indoor/outdoor) for companies processing PP fibers. In this case, the concentration level of PP particles recorded inside the company is at $26.8 \mu\text{g}/\text{m}^3$ which is higher than that recorded outside by about 2 times, respectively $13.7 \mu\text{g}/\text{m}^3$. It can be seen that considering the concentration levels of the most dangerous particles for human health, namely PM1 (PES/PET, PA, and PP).

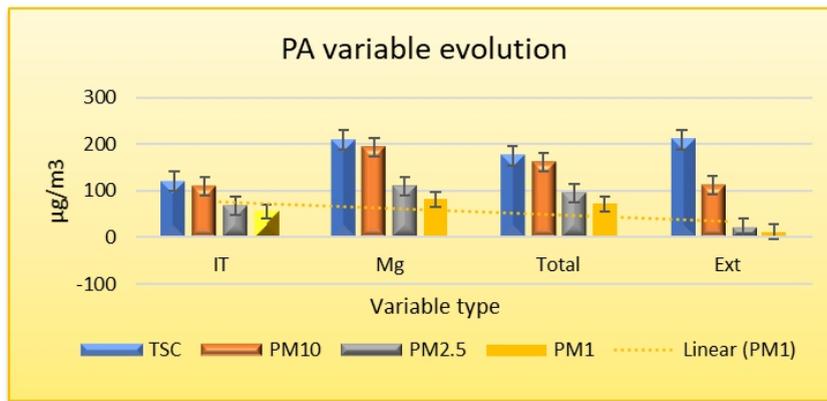


Figure 4: PA PM concentration level comparison (indoor/outdoor).

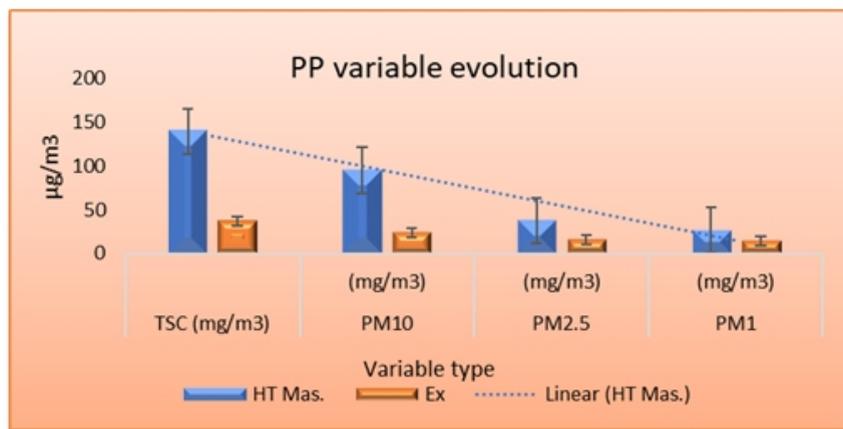


Figure 5: PP PM concentration level comparison (indoor/outdoor).

RESULTS AND DISCUSSIONS

The correlation coefficient was calculated, by using the specific equation. There is a strong correlation between the variable: "Collected mass/ Air concentration": $r = 0.997775 \div 0.999477$ and "Collected mass/ Air volume", $r = -0.97473 \div 0.80064$ for all three diameters studied. A weak correlation is between collected mass (μg) and diameter of the filters (mm): $r = 0.131679519$. The real distribution of the data series (scatter) of the analysed correlations is presented in Figure 6. The prediction of the value of the collected particle mass as a function of the diameter of the filters, particle concentration in the air, air flow of the pumps, and total air volume was made by plotting the regression curves and box-plot graphs and obtaining the regression equations.

Two types of filter packaging systems were used to transport the filters to the ringing laboratories. System A consisted of the GILAIR PLUS pump holder into which the filters were inserted and transported after weighing (see Figure 7a). The system was used to collect and transport filters of quart

ø 37mm, nucleopore ø 25 mm, and Si ø 9mm diameter, used for the collection of PES, PA and PP particles.

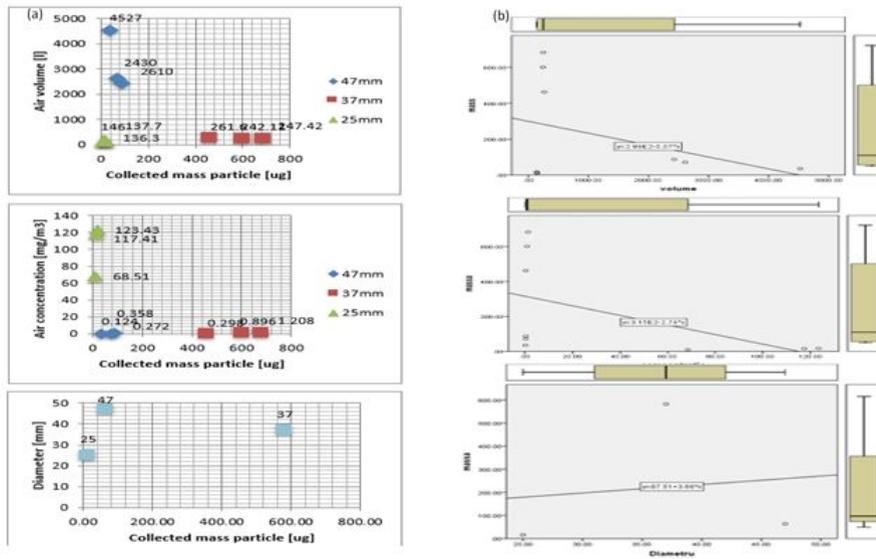


Figure 6: Correlations between air sampling parameters.

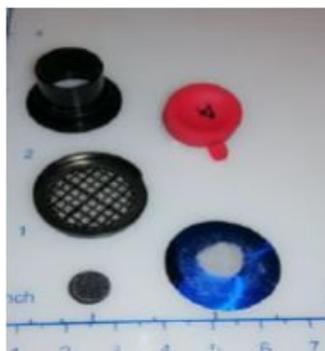


Figure 7: a) Holder pump GilAir plus for system.



Figure 7: b) Filtration components for system B.

System B is a particle filtration system consisting of 1) a metal sample holder; 2) stainless steel O-Ring; 3) a graduation cup; 4) an end plug; 5) a tube adapter; 6) a multitool (see Figure 7b). This system was used for the collection and transport of PP particles collected on 9mm diameter Si filters.

Versa 3D scanning electron microscope from Termo Scientific (formerly FEI) was used for the analysis of the microparticles. The samples were not covered with metal or carbon. The acceleration voltage used was 4 kV with a spot size between 2 and 4 (samples), the current between 0,5 pA and 12 pA, and the WD distance used was 10mm. The nucleopore sample had a circular trace with an area of about 336 mm². For the SEM analysis of the sample five different regions with a surface of 0.46 μm^2 were selected on different parts of the sample.

Figure 8 shows SEM images for PES particles. In the case of the A system of collection and transportation, very low concentrations of particles on filters were identified. A small number of particles (50) were counted. High-resolution image showing the fiber-like structure and small particles (with size below 10 microns) for all types of particles.

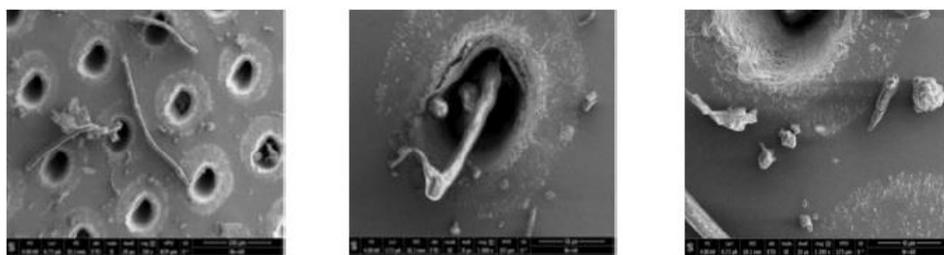


Figure 8: SEM images for PES/PET particles and microfibrils.

In the case of quartz filters, most particles would likely be deposited further inside the filter instead of on the surface, so any microscopical method will not work to determine concentrations. Particles and fibers were identified using μ Raman spectroscopy: an optical microscope image of microparticles and fiber fraction of PES, Raman spectra of Si and Raman spectra of PES, Raman mapping centered on the 1600 cm^{-1} line of the PES footprint, for PES particles collected and transported with system A are shown in Figure 9. Figure 10 shows the GS-MS spectra for the determination of the amount of PES particles.

GS-MS analyses showed us the presence of PET particles but also a small number of polystyrene particles. In the case of PP particles, a high quantity of PET particles was identified. For the PA particles, because of the low quantity of the particles, it wasn't possible to provide GS-MS analyses. To improve the particle collection procedure, system B was designed and provided for the selective collection of PM₁₀ and PM₁ particles and their analysis. The system was coupled with the Gil Air Plus pump (see Figure 11) and applied for the collection of PP particles on 9mm Si filters and will be further applied for PES particles.

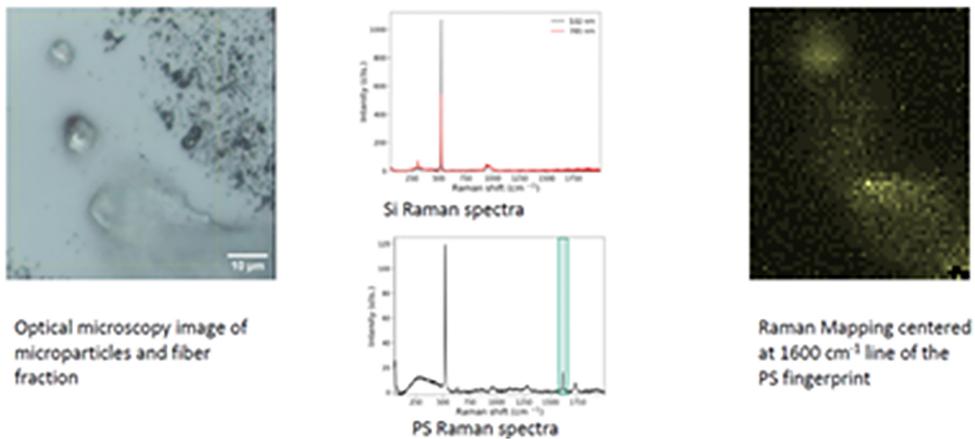


Figure 9: Optical microscope image and μ Raman spectra (PES particles).

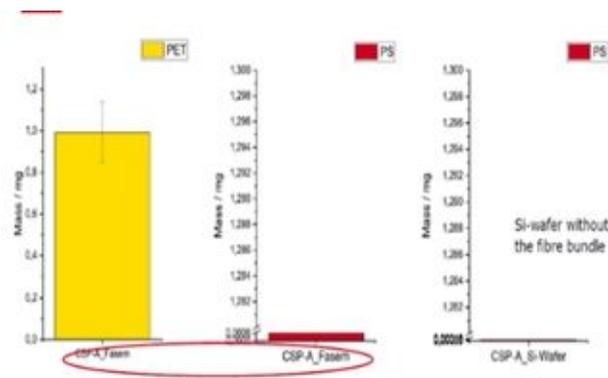


Figure 10: GS-MS - PES particles.



Figure 11: Mounting of system B.

SEM images of PM10 and PM1 PP particles are shown in Figure 12 a, b.

Both on the 10 μm and 1 μm pore size filter, different kind of particles and fibers can be detected all over the sample. As shown in Raman spectra for PM10 (see Figure 13a), and PM1 (see Figure 13b), the number of particles present on the 1 μm pore size filter is very low, compared with the particles presented on the 10 μm pore size filter.

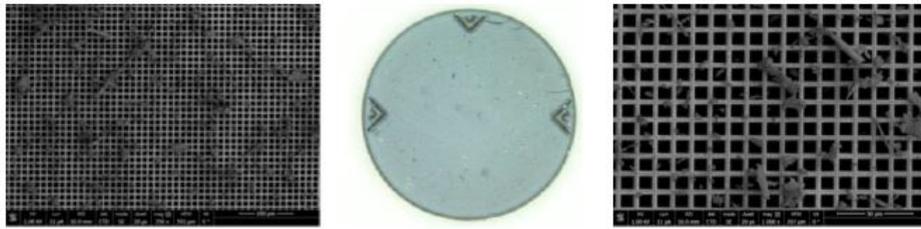


Figure 12: a) SEM - PM10.



Figure 12: b) SEM - PM1.

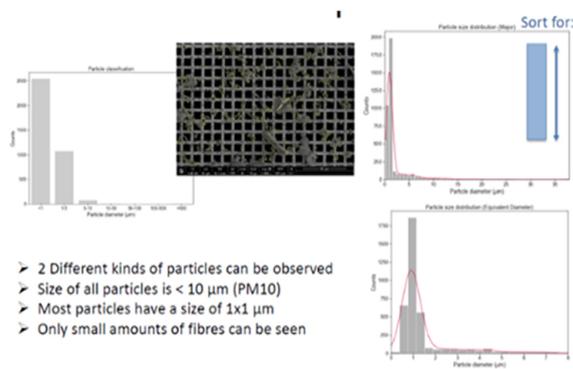


Figure 13: a) Raman spectra for PM10.

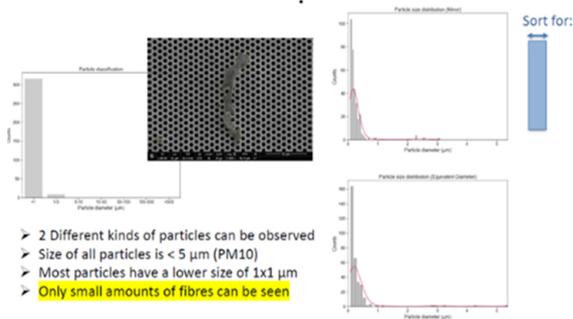


Figure 13: b) Raman spectra for PM1.

Figure 14 shows the GS-MS analyses for PM10 and PM1 which showed that the amount of 10 µm particles is about 10 times that of 1 µm particles. A small amount of polyester was also identified.

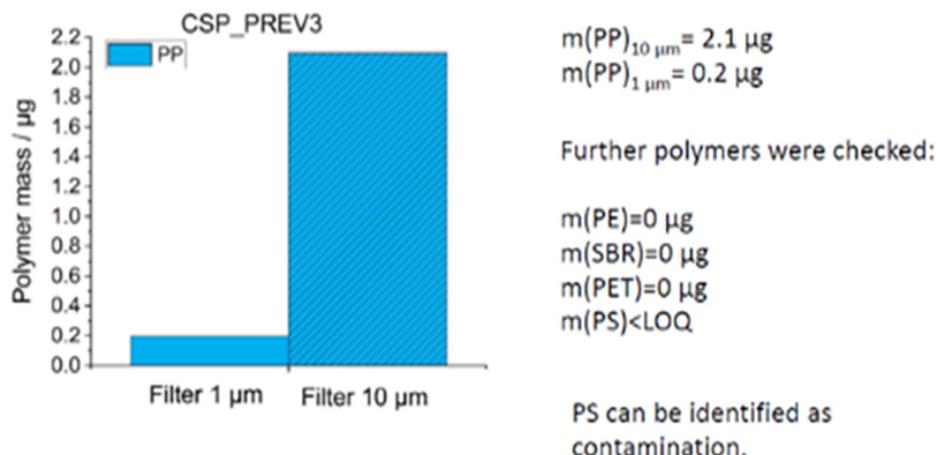


Figure 14: GS-MS for PM10 and PM1.

CONCLUSION

For all categories of PM1 concentrated particles determined inside the working rooms is higher compared to that determined outside.

Using system A, two issues were highlighted for collection and transport: quartz filters cannot be used for the analysis of particles collected by known methods; the amount of particles on the filters is insufficient for analysis, either due to the collection parameters used or due to particle losses during transport.

Using system B for the collection and transport of Si filters, sufficient quantities of PP particles were obtained for SEM, Raman and GS-MS analyses. The system will be applied also to PES particles. Another system C for collection and transport will be applied for nucleopore filters. The GS-MS detection of foreign particles requires more careful cleaning of the workplaces and the air in the workrooms when changing batches of processed textile material (composition).

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REFERENCES

- Agay-Shay, K. et al., 2013. Air pollution and congenital heart defects. *Environ. Res.*, Volume 124, pp. 28–34.
- Grimm Aerosol Technik, n.d. *Optical spectrometer 11-E*. [Online] Available at: <https://www.directindustry.com/prod/grimm-aerosol-technik/product-69071-1571949.html>
- Grosu, M. C. et al., 2023. Collection and Characterization of Synthetic Airborne Particles. *Advanced Materials Letters*, 14(1).

- Klingelhofer, D., Braun, M., Quarcoo, D. & Brüggmann, D., 2020. Research landscape of a global environmental challenge: Microplastics. *Water Research*, 170(115358), pp. 1–14.
- Qader, W., Dar, R. A. & Rashid, I., 2023. Phytolith particulate matter and its potential human and environmental effects. *Environmental Pollution*, Volume 37, p. 121541.
- Schopel, B. & Stamminger, R., 2019. A comprehensive literature study on microfibres from washing machines. *Tenside Surfactants Detergents*, 56(2), pp. 94–104.
- Visileanu, E. et al., 2023. The Structural Influence of Airborne Particles on their Toxicity. *Advanced Materials Letters*, 14(2), pp. 2302–1718.
- von Schneidemesser, E. et al., 2015. Chemistry and the Linkages between Air Quality and Climate Change. *Chemical Reviews*, 115(10), pp. 3856–3897.